



DNA Extraction and Purification

Lab Equipment

Automatic pipettes



Vortex



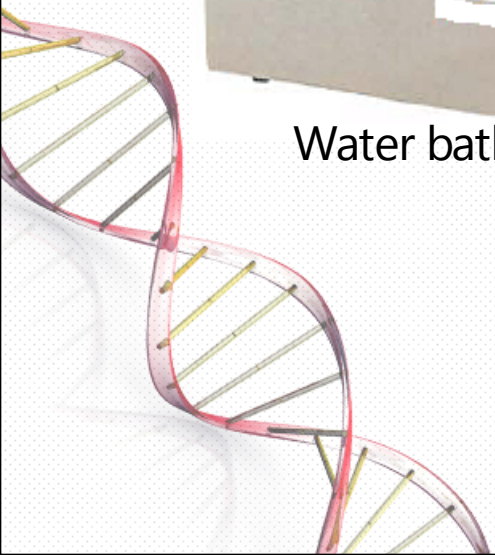
Microcentrifuge



Water bath



UV-spectrophotometer



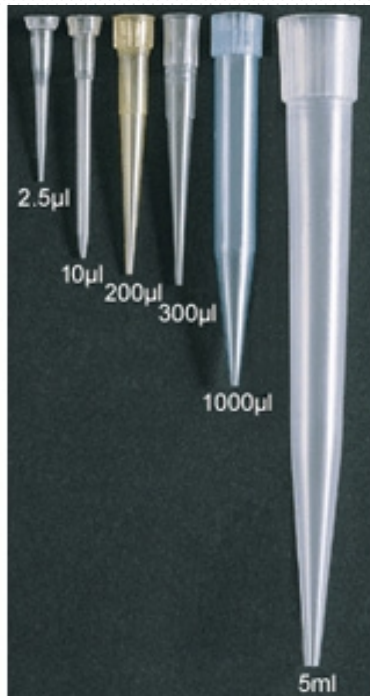
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Eppendorf tube



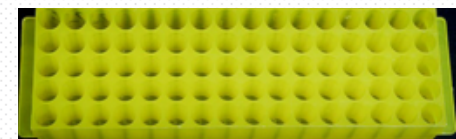
Cuvettes



Tips



Rack- test tube



Rack- eppendorf tube



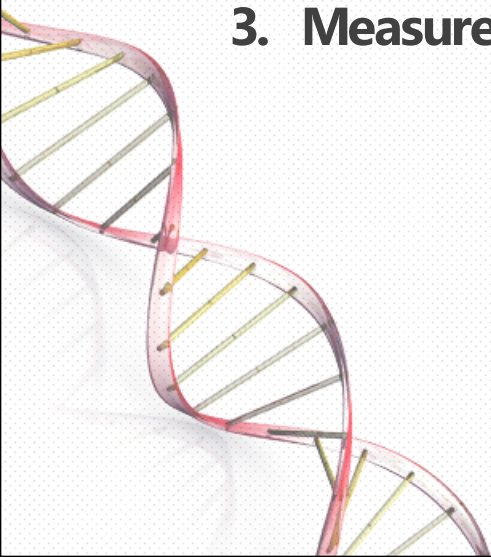
DNA Extraction

Principle:

1. **Lysis of nucleated cells.**
2. **Removal of contaminants:** Any substance other than DNA, e.g., proteins.
3. **Measurements:** UV absorbance at 260nm and 280nm

Purity of DNA solution: 260/280 ratio

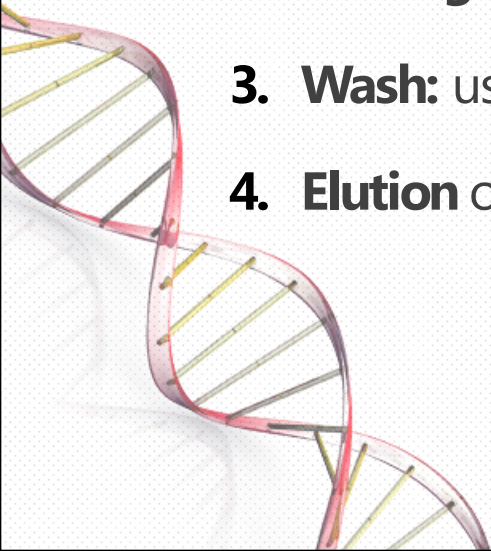
DNA concentration: Absorbance at 260nm



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Steps:

1. **Lysis** of nucleated cells using lysis buffer.
2. **Binding** of DNA to the membrane of spin column.
3. **Wash**: using wash buffer.
4. **Elution** of pure DNA.





Spin Protocol of DNA Purification from Blood

Video



<https://www.youtube.com/watch?v=gmnw6CWtN5k>



1. Pipette **20 μ l** protease.
2. Add **200 μ l** sample.
3. Add **200 μ l** Buffer **AL**.



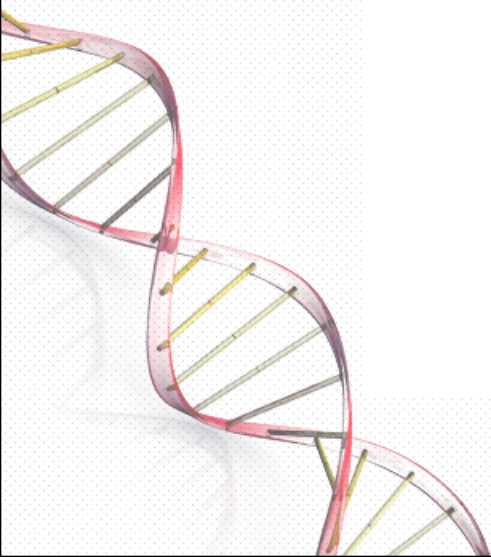
4. Mix by pulse-vortex for 15s and incubate at **56°C** for **10 minutes**.
5. Briefly centrifuge.
6. Add **200 μ l** 96-100% ethanol and mix by pulse-vortex for 15s.
7. Briefly centrifuge.



8. Apply the mixture to the Mini spin column.
9. Centrifuge at **8000 rpm** for **1 minute**.



10. Discard the collection tube.
11. Place the column in a new 2ml collection tube.
12. Add **500 μ l** buffer **AW1**.
13. Centrifuge at **8000 rpm** for **1 minute**.





14. Place the column in a new 2ml collection tube.
15. Add **500 μ l** buffer **AW2**.
16. Centrifuge at **14000 rpm** for **3 minutes**.
17. Place the column in a new 2ml collection tube.
18. Centrifuge at **14000 rpm** for **1 minute**.

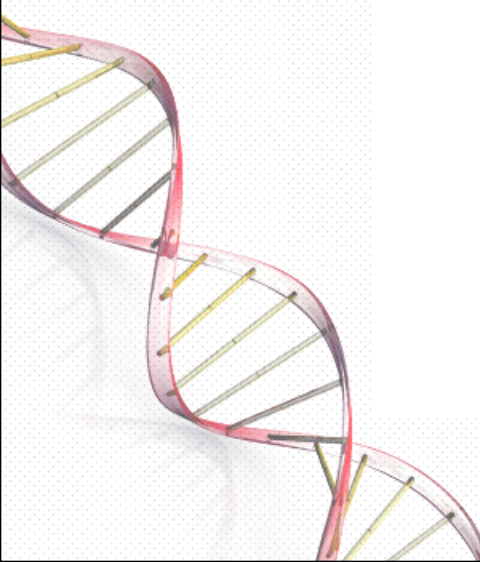


19. Place the column in a new 1.5ml tube.
20. Add **200 μ l** buffer **AE**.
21. Incubate at room temperature for **1 minute**.
22. Centrifuge at **8000 rpm** for **1 minute**.



Add 400 μ l buffer AE to dilute the eluted DNA (3X dilution)

23. Quantify the DNA concentration.





Quantification of the purified DNA

Measurements

- Done by
the spectrophotometer
- Measure the Absorbance at **260nm** and at **280nm**.
 - **Assess the DNA purity:** 260/280 ratio
(Accepted ratio: **1.7 - 1.9**)
 - **Calculate DNA Conc.:** Provided $A_{260} = 1.0$, DNA is $50 \mu\text{g/ml}$, unknown DNA Conc. can be calculated by cross multiplication

$$A_{260} = 1.0$$

$$\text{DNA conc.} = 50 \mu\text{g/ml}$$

$$A_{260} = 0.5$$

$$\text{DNA conc. ?}$$

Note: In case of diluting the eluted sample, multiplies the final concentration by the dilution factor. This can be adjusted by the spectrophotometer.

DNA Yield

DNA Yield = DNA Volume x final DNA Conc.

Example:

If you have

Volume of DNA solution: 200 μ l (0.2 ml)

Final DNA Conc.: 30 μ g/ml

**Then, the yield (μ g) = 0.2 ml x 30 μ g/ml
= 6.0 μ g**

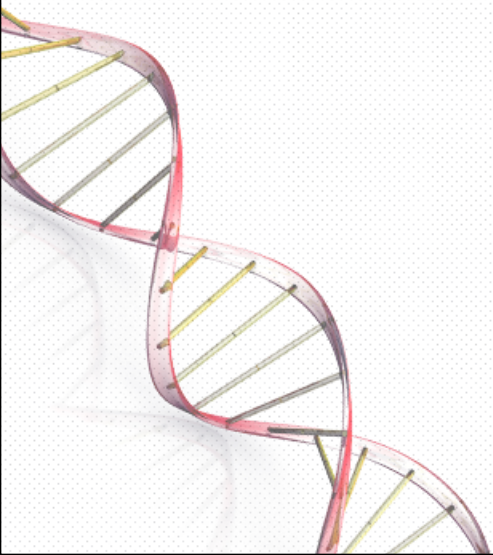




Molecular Techniques and Applications

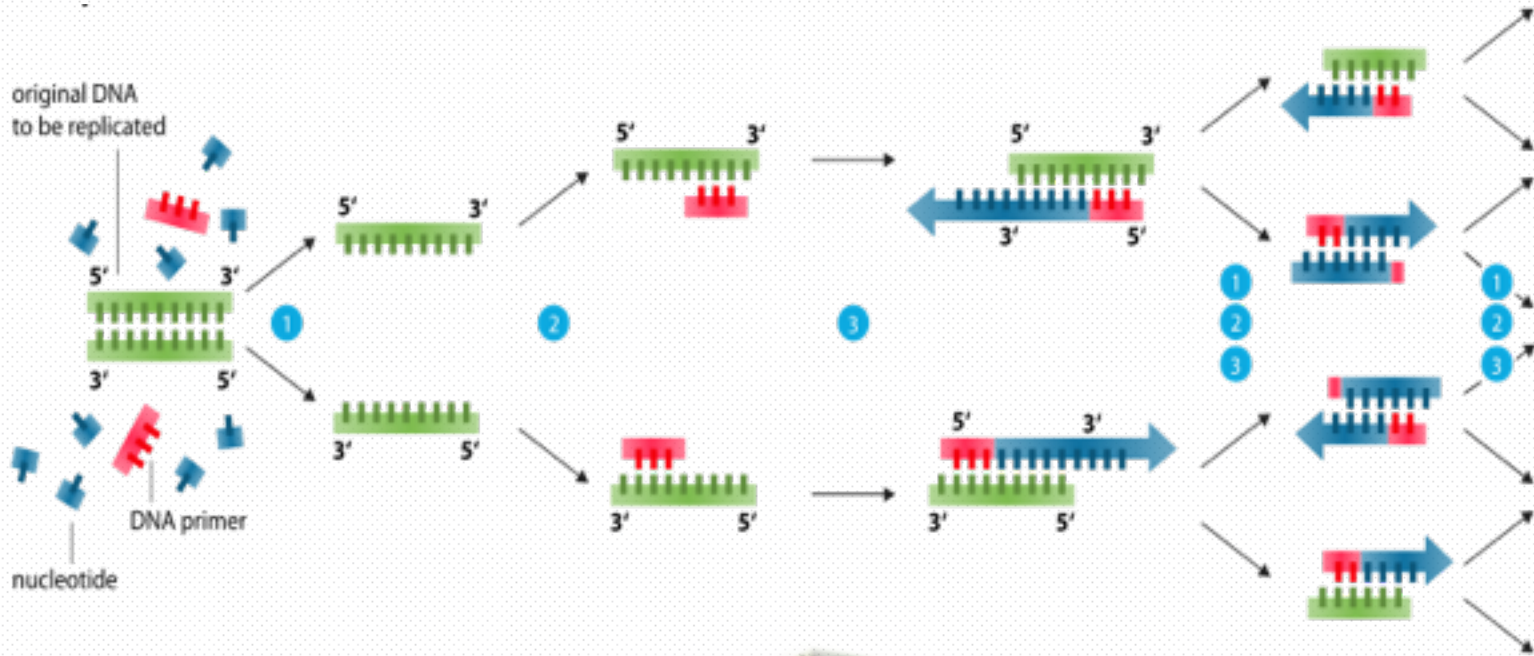
Note:

Almost all molecular biology techniques can be utilized for diagnosis and research



a. Amplification techniques:

e.g. Polymerase Chain Reaction (PCR)



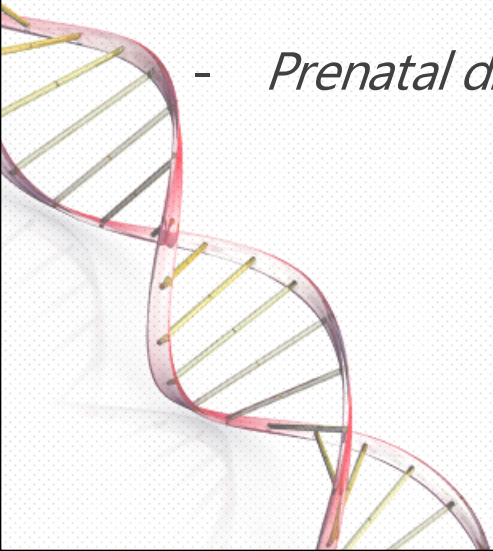
- 1 Denaturation at 94-96°C
- 2 Annealing at ~68°C
- 3 Elongation at ca. 72 °C



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Applications of PCR:

- *Comparison of a normal gene with a mutant form of the gene.*
- *Detection of low-abundance nucleic acid sequences.*
- *Forensic analysis of DNA samples.*
- *Prenatal diagnosis.*



Other examples of molecular techniques:

1. Restriction Fragment length polymorphism (RFLP).



<https://www.ncbi.nlm.nih.gov/probe/docs/techrflp/>

2. Southern blotting.



<http://www.onlinebiologynotes.com/southern-blotting-principle-procedure-application/>

